

# (-)-CRYPTOCARYALACTONE AND (-)-DEACETYLCRYPTOCARYALACTONE—GERMINATION INHIBITORS FROM *CRYPTOCARYA MOSCHATA* SEEDS

GAYLAND F SPENCER, ROGER E ENGLAND and REBECCA B WOLF

Northern Regional Research Center, Agricultural Research Service, U S Department of Agriculture\*, Peoria, Illinois 61604, U S A

(Received 13 March 1984)

**Key Word Index**—*Cryptocarya moschata*, Lauraceae, 2-pyranones, seed germination inhibitor

**Abstract**—(-)-Cryptocaryalactone (6-[2-acetoxy-4-phenyl-3-butenyl]-5,6-dihydro-2-pyranone) and (-)-deacetylcryptocaryalactone (6-[2-hydroxy-4-phenyl-3-butenyl]-5,6-dihydro-2-pyranone) isolated from *Cryptocarya moschata* seeds are natural germination inhibitors. Applied at 0.004 M, the second compound completely arrested germination of velvetleaf (*Abutilon theophrasti*) and decreased the germination rate of soybeans, but did not appear to affect corn. The first compound was not as effective, 0.004 M reduced velvetleaf germination 50%.

## INTRODUCTION

Substituted 2-pyranones are important natural products with diverse biological activities [1]. Reported germination inhibitors include parasorbic acid [2] and psilotin [3]. In the course of our effort to find compounds with weed control potential [4], extracts of *Cryptocarya moschata* Nees and Mort showed good activity against velvetleaf (*Abutilon theophrasti* Medic), a troublesome competitor of corn and soybeans. Isolation and characterization of the active principles proved them to be (-)-cryptocaryalactone (1) and its deacetyl analog (2) (+)-Cryptocaryalactone was isolated from *C. bourdillonii* [5] and 6-(2-phenylethenyl)-5,6-dihydro-2-pyranone from *C. caloneura* [6], but this is the first report of levorotatory cryptocaryalactone, of its deacetyl analog, and of their antigermination activity.

## RESULTS AND DISCUSSION

Compound 1 gave mass and NMR spectra identical to those of (+)-cryptocaryalactone [5]. These spectra were also exhibited by the acetyl derivative of compound 2. Both were levorotatory, the magnitude of rotation for 1 (-20°) is about the same as that for (+)-cryptocaryalactone (+16°) [5].

The respective antigermination activities of the two are given in Table 1. Compound 2 was decidedly more active against velvetleaf, so further experiments to determine its toxicity against the crop plants of corn and soybeans were performed. Corn was relatively unaffected at a concentration virtually lethal to velvetleaf (0.004 M) but soybean germination was about 75% as great as controls. These compounds are not as toxic as benzyl isothiocyanate [4] but have an effect similar to that of psilotin on turnips [3].

The activity of psilotin could be reversed by the

Table 1 Effect of cryptocaryalactones 1 and 2 on germination

Compound and concentration (M)	% Germination relative to controls		
	Velvetleaf	Soybeans	Corn
1 0.004	55*	—	—
1 0.001	94	—	—
2 0.004	6*	79*	83
2 0.002	59*	92	100
2 0.001	59*	105	90
2 0.0008	85		
2 0.004 + 0.0003 GA <sub>3</sub> † simultaneous treatment	15*		
2 0.004 + 0.0003 GA <sub>3</sub> delayed treatment			
(1) before GA <sub>3</sub>	8*		
(2) after GA <sub>3</sub>	10*		

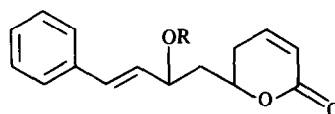
\*Significantly different from controls at 95% level or better as determined by the Chi-square one-tailed test.

†Gibberellic acid

addition of gibberellic acid (GA<sub>3</sub>) to the medium [3]. Our experiments (Table 1) showed that GA<sub>3</sub> had no effect on the activity of compound 2 applied to velvetleaf.

## EXPERIMENTAL

The *C. moschata* seeds were collected in Uruguay and were identified by botanists at the Beltsville Agricultural Research Center, Beltsville, MD. Seeds (150 g) were separated into kernel



R  
 1 Ac  
 2 H

\*The mention of firm names or trade products does not imply that they are endorsed or recommended by the U S Department of Agriculture over other firms or similar products not mentioned.

(50 g) and hull (100 g) fractions. Each fraction was then finely ground and extracted in a Soxhlet with hexane (8 hr) and  $\text{Me}_2\text{CO}$  (16 hr). The two extracts of the hulls were combined, slurried onto a dry silica column, and eluted sequentially with 300 ml portions of EtOAc-hexane (1:9), EtOAc-hexane (1:1), EtOAc-hexane-EtOH (25:25:1) and EtOAc-hexane-EtOH (5:5:1). HPLC of the fractions on Partisil 10/50 PAC (Whatman) with EtOAc-hexane-EtOH (66:33:5) gave highly enriched portions of 1 and 2. These were purified by reversed-phase HPLC on a Zorbax C-8 (Dupont) column eluted with  $\text{Me}_2\text{CO}-\text{H}_2\text{O}$  (1:1). Bioassays were used as a guide to active materials throughout the isolation procedure.

The hexane extract of seed kernels produced crystals after most of the hexane had been removed. These crystals were recovered and washed  $\times 3$  with cold hexane. TLC on silica plates [0.25 mm, Brinkman,  $\text{C}_6\text{H}_6$ -EtOH (9:1)] showed them to be virtually pure 1. The remainder of this extract and the  $\text{Me}_2\text{CO}$  extract were taken through the chromatographic steps given above for the hull extracts. Total yields from all processes were 1, 137 mg and 2, 286 mg. Optical rotations were measured in  $\text{CHCl}_3$ , 1,  $[\alpha]_{\text{D}}^{27} -20^\circ$  (c 1.0), 2,  $[\alpha]_{\text{D}}^{27} -94^\circ$  (c 2.2).

Compound 2 was allowed to stand overnight in  $\text{Ac}_2\text{O}$ -pyridine (1:2) and the product (1) was recovered by ether/ $\text{H}_2\text{O}$  extraction.

Mass and NMR spectroscopic equipment and techniques and details of the bioassay procedure are described in previous work [4, 7]. Concentrations are given in Table 1. To test whether or not the activity could be negated or reversed by gibberellic acid,  $\text{GA}_3$  and compound 2 were applied together (simultaneous treatment, Table 1), or  $\text{GA}_3$  was added 2 days after the original treatment with 2 and another 2 days were allowed to elapse before germination was evaluated (delayed treatment).

## REFERENCES

- 1 Adityachaudhury, N and Das, A K (1979) *J Sci Ind Res* **38**, 265
- 2 Haynes, L J and Jones, E R H (1946) *J Chem. Soc* 954
- 3 Siegel, S M (1976) *Phytochemistry* **15**, 566
- 4 Wolf, R B, Spencer, G F and Kwolek, W F (1984) *Weed Sci* (in press)
- 5 Govindachari, T R, Parthasarathy, P C and Modi, J D (1972) *Indian J Chem* **10**, 149
- 6 Hlubucek, J R and Robertson, A V (1967) *Aust J Chem* **20**, 2199
- 7 Spencer, G F and Flippen-Anderson, J L (1981) *Phytochemistry* **20**, 2757